

SPECIFICATION: AmberGen Miralys™ Protocol Starter Kit – Immunology Version

PURPOSE

The Starter Kit was created to provide researchers with many of the materials needed to establish the Miralys™ Laboratory Workflow in their labs. Additionally, practice slides and probe mix enable initial Miralys™ runs without using up precious samples.

This kit is to be used in conjunction with MALDI HIPLEX-IHC MIRALYS™ IMAGING LABORATORY WORKFLOW document ("Miralys™ Protocol") which has been provided. If you do not have a copy, please contact support@ambergen.com to obtain one.

CONTENTS

- 1. One (1) 6-plex Miralys[™] probe mixture store at -20C and protect from prolonged light exposure. *Details in table below*.
- 2. Set of FFPE human tonsil slides store at 4C or at room temperature:
 - a. One (1) "pre-stained" slide stained with the same 6-plex Miralys™ probe mixture as detailed in item 1
 - b. Two (2) "unstained" slides sourced from the same tissue block as item (a)
- One (1) multi-channel digital timer for keeping time while working through the Miralys™
 Protocol
- 4. One (1) hydrophobic PAP barrier pen used in Step 8 of the Miralys™ Protocol
- 5. Four (4) spin filter units (0.45 μm) used in Step 10 of the Miralys™ Protocol
- 6. One (1) UV Lightbox with US and European power cords used in Step 16 of the Miralys™ Protocol
- 7. Two (2) pieces of filter paper used in Step 18 of the Miralys™ Protocol
- 8. One (1) glass petri dish (bottom only) used in Step 18 of the Miralys™ Protocol
- 9. One (1) plastic petri dish top only used in Step 18 of the Miralys™ Protocol
- 10. Four (4) neodymium magnets used in Step 18 of the Miralys™ Protocol

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PROBE MIX DETAILS

The Miralys™ probe mixture included with the Kit has also been used to stain the pre-stained slide.

Antibody	PC-MT (Da)*	Host	Species Reactivity	Final Concentration
CD3ε	1,161.65	Rabbit	Н	2.5 μg/mL
CD68	1,216.75	Rabbit	Н	2.5 μg/mL
Collagen-1A1	1,234.87	Rabbit	M, H	2.5 μg/mL
Ki67	1,320.76	Mouse	Н	2.5 μg/mL
PanCK	1,628.78	Rabbit	M, R, H, Mk	2.5 μg/mL
VIM (Vimentin)	1,230.84	Rabbit	M, R, H, Mk	2.5 μg/mL

^{*}PC-MT (Da) = Monoisotopic (M+H)⁺ of the mass reporter

INSTRUCTIONS FOR USE

Image the Pre-Stained Slide

- Open the pink bubble-wrap slide envelope and the vacuum-sealed package within. Remove the
 pre-stained slide and re-store the unstained slides. Important note: If slides were stored cold,
 equilibrate to room temperature before opening the vacuum-sealed pouch.
- 2. Begin at Step 16 of the Miralys™ Protocol (photocleavage) and follow the protocol through to the end to prepare slide
- 3. Image in any MSI instrument

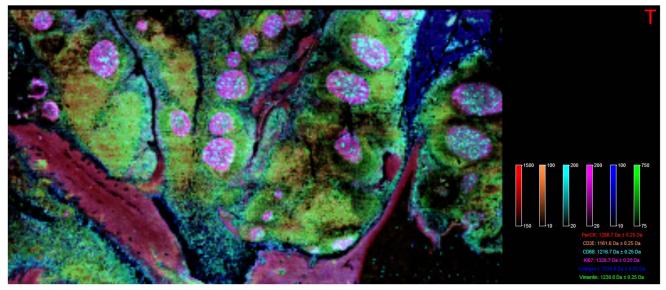
Stain and Image the Unstained Slides

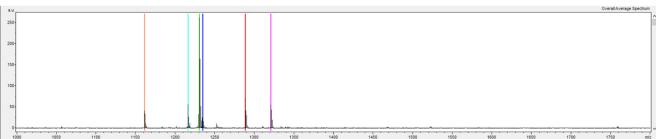
- 1. To prepare the unstained slides, begin by pre-melting as per Step 1 of the Miralys™ Protocol
- 2. Complete Steps 2, 4, and 5 of the Miralys^{\dagger} Protocol. Step 3 can be omitted because the tissue is FFPE.
- 3. Follow Steps 6 thru 8 of the Miralys™ Protocol, using alkaline antigen retrieval buffer in Step 6
- 4. Because the Miralys[™] probe is pre-mixed, perform the following in place of Step 9:
 - a. Prior to opening probe:
 - i. Vortex for 30 seconds with a benchtop vortex
 - ii. Centrifuge for 1 minute at full speed
 - b. For each slide, aliquot 24 μ L of the supplied 48 μ L probe mixture into a separate microcentrifuge tube and dilute to 400 μ L final volume with the Tissue Blocking Buffer. Vortex for 30 seconds and centrifuge again for 1 minute.
- 5. Begin again with the Miralys™ Protocol at Step 10 and follow through to the end to prepare slides.
- 6. Image in any MSI instrument

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Example Results:





Top: Example Miralys™ Results from Pre-Stained FFPE Human Tonsil Tissue Slide. MALDI mass spectrometry imaging was performed on a Bruker rapifleX™ Tissuetyper with 20 μm spatial resolution. The image was generated from flexImaging with TIC normalization and the display intensity scale in absolute units.

Bottom: Mean Spectrum from the Entire Region Shown. Color-coded bars indicate the reporter peaks from the photocleaved mass-tags (bars are color-coded according to the key provided in the image in the top panel).

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