

# Miralys™ Protocol Starter Kit: Neurology Version

### **Purpose**

The Starter Kit was created to provide researchers with many of the materials needed to establish the Miralys™ Laboratory Workflow in their labs. Additionally, practice slides and probe mix enable initial Miralys™ runs without using up precious samples. This kit is to be used in conjunction with MALDI HiPLEX-IHC MIRALYS™ IMAGING LABORATORY WORKFLOW document ("Miralys™ Protocol") which has been provided. If you do not have a copy, please contact support@ambergen.com to obtain one.

### Contents

- 1. One (1) 6-plex Miralys<sup>™</sup> probe mixture store at -20°C and protect from prolonged light exposure. Details in table below.
- 2. Set of FFPE mouse brain slides store at 4°C or at room temperature
  - a. One (1) "pre-stained" slide stained with the same 6-plex Miralys™ probe mixture as detailed in item 1
  - b. Two (2) "unstained" slides sourced from the same tissue block as item (a)
- 3. One (1) multi-channel digital timer for keeping time while working through the Miralys™ Protocol
- 4. One (1) hydrophobic PAP barrier pen used in Step 8 of the Miralys™ Protocol
- 5. Four (4) spin filter units (0.45 μm) used in Step 10 of the Miralys™ Protocol
- 6. One (1) UV Lightbox with US and European power cords used in Step 16 of the Miralys™ Protocol
- 7. Two (2) pieces of filter paper used in Step 18 of the Miralys™ Protocol
- 8. One (1) glass petri dish (bottom only) used in Step 18 of the Miralys™ Protocol
- 9. One (1) plastic petri dish top used in Step 18 of the Miralys™ Protocol
- 10. Four (4) neodymium magnets used in Step 18 of the Miralys™ Protocol



### **Probe Mix Details**

Target	PC-MT (Da)*	Host	Reactivity	Final Concentration
GFAP	1011.55	Rabbit	M, R, H	2.5 μg / mL
GLUT1 (SLC2A1)	856.56	Rabbit	M, R, H, Mk	2.5 μg / mL
MBP (Myelin Basic Protein)	1365.73	Rabbit	M, R, H	1.25 μg / mL
NeuN	1308.71	Rabbit	M, R, H	2.5 μg / mL
NF-L (Neurofilament Light)	1345.74	Rabbit	M, R, H	1.25 μg / mL
SYN-I (Synapsin I)	1482.77	Rabbit	M, R, H	1.25 μg / mL

<sup>\*</sup>PC-MT (Da) = Monoisotopic (M+H)+ of the mass reporter

### **Instructions for Use**

### **Image the Pre-Stained Slide**

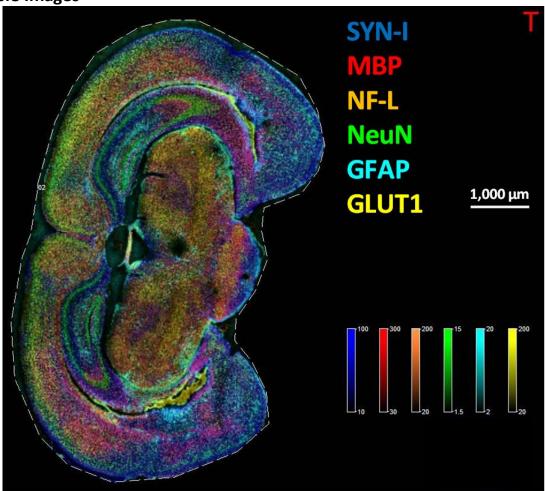
- Open the pink bubble-wrap slide envelope and the vacuum-sealed package within. Remove the pre-stained slide and re-store the unstained slides. Important note: If slides were stored cold, equilibrate to room temperature before opening the vacuum-sealed pouch.
- 2. Begin at Step 16 of the Miralys™ Protocol (photocleavage) and follow the protocol through to the end to prepare slide.
- 3. Image in any MSI instrument.

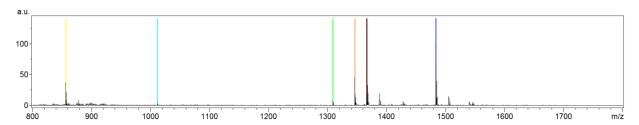
# Stain and Image the Unstained Slides

- 1. To prepare the unstained slides, begin by pre-melting as per Step 1 of the Miralys™ Protocol.
- 2. Complete Steps 2, 4, and 5 of the Miralys™ Protocol. Step 3 can be omitted because the tissue is FFPE.
- 3. Follow Steps 6 thru 8 of the Miralys™ Protocol, using citrate antigen retrieval buffer in Step 6.
- Because the Miralys™ probe is pre-mixed, perform the following in place of Step 9:
  - a. Prior to opening probe:
    - i. Vortex for 30 seconds with a benchtop vortex
    - ii. Centrifuge for 1 minute at full speed
  - b. For each slide, aliquot 24 μL of the supplied 48 μL probe mixture into a separate microcentrifuge tube and dilute to 400 µL final volume with the Tissue Blocking Buffer. Vortex for 30 seconds and centrifuge again for 1
- 5. Begin again with the Miralys™ Protocol at Step 10 and follow through to the end to prepare slides.
- 6. Image in any MSI instrument.



## **Sample Images**





**Top:** Example Miralys™ Results from Pre-Stained FFPE mouse brain tissue slide. MALDI mass spectrometry imaging was performed on a Bruker rapifleX™ Tissuetyper with 20 μm spatial resolution. The image was generated from flexImaging with TIC normalization and the display intensity scale in absolute units.

Bottom: Mean Spectrum from the Entire Region Shown. Color-coded bars indicate the reporter peaks from the photocleaved mass-tags (bars are color-coded according to the key provided in the image in the top panel).